



# Inter-prismatic matrix structure characterization of mollusk shell and its effect on crystal formation

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## Abstract

Mollusk biomineralization is an elaborate process in which cells, organic macromolecules, and calcium carbonate crystals are actively involved. Macromolecules (mainly are proteins and polysaccharide) act as a key role in regulating and limiting the size, orientation, polymorph and texture of inorganic phase. In this work, we focused on the inter-prismatic matrix of mollusk shell combining scanning electron microscopy (SEM), X-ray diffraction (XRD) and transmission electron microscopy (TEM) analytical techniques with  $\text{CaCO}_3$  recrystallization experiment to characterize its structure and effects on crystal formation. Our results show that the inter-prismatic matrix is not a sort of pure polymer, calcite nano-crystals are also located inside the inter-prismatic matrix. Interestingly, it seems that these nanocrystals have a preferred orientation, which means the inter-prismatic matrix do impose effect on the crystal formation. In vitro re-crystallization experiment using partially dissolved prismatic fragment as template indicates that the (104) faces of  $\text{CaCO}_3$  micro-crystals are closely associated with the walls of inter-prismatic matrix. Furthermore, a possible growth mechanism of mollusk shell prismatic layer was proposed.

**Key Words:** Biomineralization, Mollusk shell, Inter-prismatic matrix, Calcite, Dissolution

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## 1. Introduction

Billions of years' evolution endows living organism with fascinating shapes, structures and corresponding multi-function features which are attracting more and more scientists to throw themselves into this field called biomineralization [1,2]. Of all the minerals formed by organisms, calcium carbonate ( $\text{CaCO}_3$ ) is well known not only because it is widely distributed in nature, but also because it is a kind of typical mineral to reveal the biomineralization mechanism which is mystical to the curious people. On the other hand, understanding the design strategy behind the mineral will hold great promise for the future development of biological, chemical and materials sciences [3-9].

Mollusk is a kind of classic model organism for the research of biomineralization [4]. Most mollusk secretes  $\text{CaCO}_3$  crystals in the form of aragonite and/or calcite

which are well studied using modern materials analysis and characterization techniques, such as X-ray diffraction (XRD) [10], scanning electron microscope (SEM) [11,12], transmission electron microscopy (TEM) [13], and atomic force microscopy (AFM) [14], etc. Based on these advanced analytical techniques, some hypotheses [15-23] are also put forward in attempt to reveal the formation mechanism of mollusk shell. However, the shell is a special composite which includes two parts: inorganic phase and organic phase. Deduced alone from the perspective of inorganic phase, the authentic scenario is less far revealed. To fully understand the mollusk shell mineralization process, an organic phase view is also equally required. The extracellular matrix (organic phase) is made up of multifunctional macromolecules (mainly polysaccharides and proteins) [2] which play a key role

in regulating and controlling  $\text{CaCO}_3$  crystals growth and orientation, determining the shape, size and polymorph [24-29], maintaining the mechanical performances [18], even displaying enzymatic functions [30] and involving in cell signalling [31]. Currently, there is a great deal of work relating with the purification and characterization of proteins from mollusk shell [32-38]. However, less concern is paid attention to the inter-prismatic proteins (insoluble in water) as a whole from a structural perspective. In this work, we focus on the insoluble inter-prismatic matrix extracted from the prismatic layer in *Atrina pectinata*, characterising its structures and certain habits through SEM, XRD, TME and  $\text{CaCO}_3$  re-crystallization experiment in vitro. In the end, we discussed the possible effects of inter-prismatic matrix on mollusk shell formation.

## 2. Materials and methods

### 2.1 Chemicals

Sodium hydroxide (A.C.S. reagent, Aldrich), hydrochloric acid (A.C.S. reagent, Aldrich), Ethanol (anhydrous, Aldrich), Calcium chloride (A.C.S. reagent, Sigma-Aldrich), ammonium carbonate (A.C.S. reagent, Aldrich) and Ethylene Diamine Tetraacetic Acid (A.C.S. reagent, Aldrich) were used without further purification. De-ionized water was used throughout all the experiments.

### 2.2 Mollusc shell collection and Treatment

The studied shell of Mediterranean fan mussel (*Atrina pectinata*) was bought alive from local seafood market (Tongchuan Road, Shanghai). Nacre and prism layers were separated mechanically. In our studies, only the prismatic layer was used for the following experiments. The surface of prismatic layer was slightly etched with  $1 \text{ molL}^{-1}$  NaOH solution (to remove debris), carefully cleaned using DI water, and then cut into several small fragments. These fragments were subsequently separated into different parts for crystal dissolution and growth experiments.

### 2.3 Decalcified with HCl and FAM

The partially- and whole-decalcified prismatic samples were prepared as follows: two pieces (1 cm x 1 cm) of previously weighed prismatic fragments (3.237 and 3.265 g) were simultaneously submerged into 10 ml  $1 \text{ molL}^{-1}$  HCl aqueous solutions, respectively. After vigorous reactions for 1 hour, one of them was taken out from the solution for drying. The remaining one was proceeded to react until 24 hours, took out from HCl solution and dried at room temperature. In contrast, another prism sample (3.243 g) was decalcified in 5% EDTA aqueous solution for 3 days. Prior to the observation of scanning electron microscopy, all the samples were sputtered with gold (2 nm).

### 2.4 $\text{CaCO}_3$ Crystal Growth by inter- prismatic matrix as template

A piece of partially-dissolved prismatic fragment

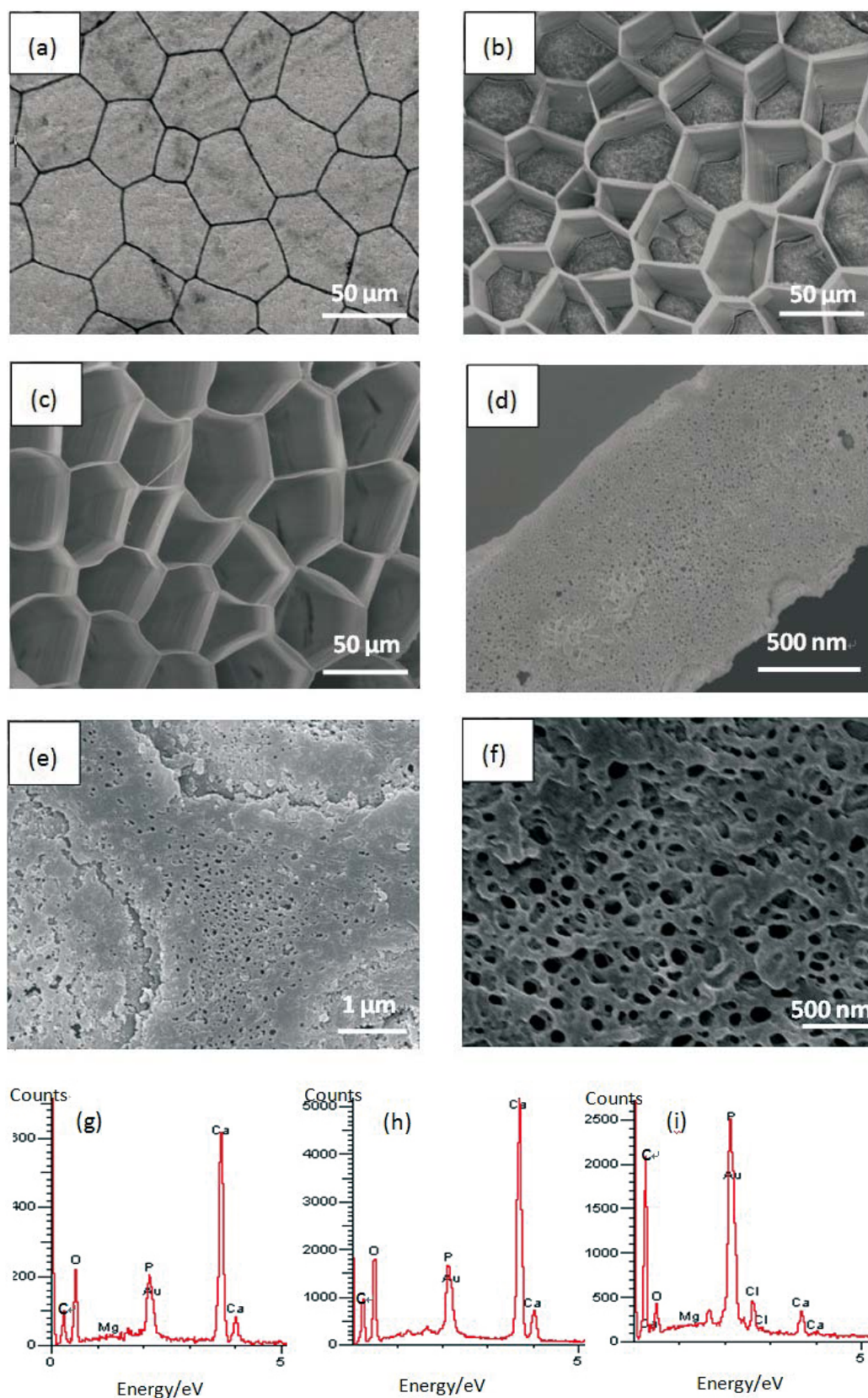
( $1 \text{ molL}^{-1}$  HCl, 30 min) was put flatly in a glass Petri dish and immersed in aqueous solutions of calcium chloride ( $[\text{Ca}^{2+}] = 10 \text{ mmolL}^{-1}$ ). The Petri dish was then placed into a sealed desiccator containing a 10 ml vial of ammonium carbonate powder.  $\text{CaCO}_3$  crystallization was induced by the slow diffusion of ammonium carbonate into calcium chloride solutions at room temperature for 6 hours. The resulting product was taken out followed by thorough washing with DI water and ethanol sequentially, and dried at room temperature before characterization.

### 2.5 Instruments for characterization

SEM/EDX observation: The morphology observation and qualitative energy dispersive X-ray (EDX) tests were performed using scanning electron microscopy (SEM/EDX, JEOL JSM-6700). Gold-sputtered, the qualitative EDX spectra of original prismatic shell and inter-prismatic matrix were recorded using an Edax (Leo 1540 XB) EDX detector mounted on the scanning electron microscope. EDX measurements were performed at 10 kV, with a probe current of  $20 \mu\text{A}$ . X-ray diffraction patterns of prism samples (powder, prismatic fragment and dissolved obtained polymer) were recorded by using a Bruker AXS GADDS X-ray diffractometer. TEM (HRTEM) imaging was done with a JEOL 2010-F TEM. The TEM foil was obtained using focused ion beam (FIB, Hitachi FE-2100) milling technique by cut perpendicularly to the inter-prismatic matrix. Before FIB cutting, the prismatic fragment was sputtered with gold.

## 3. Results and discussion

Morphology observation is performed with different method treated samples. Figure 1a. shows the SEM image of the top-surface of original prism shell. The topology is made up of many irregular polygonal unite cells which are separated by inter-prismatic matrix. The average diameter of individual unite cell is about  $30 \mu\text{m}$ , meanwhile the thickness of the wall of inter-prismatic matrix is about  $2 \mu\text{m}$ . After 30 minutes etching with HCl aqueous solution, some well-like structures are shown in Figure 1b. The depth of the well depends on the etching time. After complete dissolution, a honeycomb-like structure is shown in Figure 1c. One could see the layer by layer structures from the lateral walls, which indicate the periodic rhythmic deposition process of organic matrix [39]. Figure 1d is the higher magnification of Figure 1c, where some porous structures appear on the inter-prismatic wall. It is noted that there are no porous structures on original inter-prismatic matrix without any treatment. The same case also occurred through the use of 5% EDTA aqueous solution in figure 1e and 1f. Interestingly, both the inorganic phase and organic phase show the porous structures, which indicates that they might be calcium-containing materials (Note: EDTA is a strong Ca ions chelator). Figure 1f is the magnified picture of inter-prismatic matrix in figure 1e. The size of porous structure ranges from 50 to 100 nm. Representative EDX spectra for organic phase, inorganic phase of the original



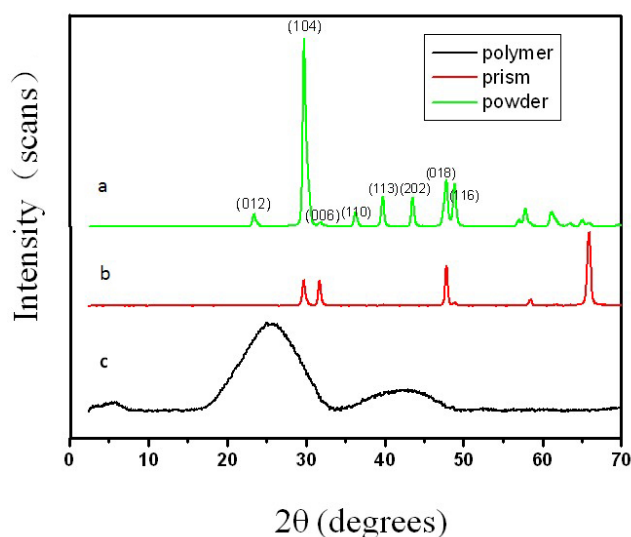
**Figure 1** SEM images of (a) original prism top surface showing a polygonal structure, the width between the boundary of two neighbouring prism-like crystals are about 2  $\mu\text{m}$ . (b) partially dissolved prism surface by  $1\text{mol}\cdot\text{L}^{-1}$  HCl aqueous solution with the arrays of well-like shape. The layered structure is clearly seen from the side walls of the wells. The depth of the well could be controlled by time. (c) the completely-dissolved honeycomb-like inter-prismatic matrix. (d) a higher amplification of the top surface of inter-prismatic matrix wall revealing the porous structures which are formed by acidic dissolution. (e) partially dissolved prism surface using 5% EDTA aqueous solution. The nano-porous structures are observed on both the inorganic and organic phase. (f) a higher magnification of top surface of inter-prismatic matrix in Figure (e). EDX results of (f) inter-prismatic matrix of original prism surface; (g) the inorganic phase of original prism surface; (h) dissolution-obtained inter-prismatic matrix. Ca signals are present in the inter-prismatic matrix. The relative intensity of Ca signals is  $h > g > i$ .

prism and dissolution obtained polymer are shown in Figure 1g-i. They exhibited strong Ca peaks as well as C and O peaks, and can be distinguished by the relative intensity. The inorganic phase shows the most intense Ca signals, which attribute to a great number of calcite crystals. It is interesting to note that the inter-prismatic matrix also presents Ca signals, which reduces in the HCl-attacked obtained polymer (The relative value of Ca is  $h>g>i$ ). Based on these, we speculated that  $\text{CaCO}_3$  polymorph (crystalline or amorphous) might be included in the inter-prismatic matrix. Although Mg as an important element is often involved in the formation of biogenic  $\text{CaCO}_3$  [40], we only observe the neglectable amount of Mg signal in our species. Additionally, all Au signal present in the spectra is the result of sputtering.

X-ray diffraction patterns were then measured from the ground shell powder, original prism outer shell and completely dissolved prism surface. The powder X-ray diffraction pattern (Figure 2a) indicated that the mineral phase is predominantly calcite with rhombohedral crystallographic unit cell with parameters  $a=b=4.990$  and  $c=17.061$  Å. The signal corresponding to  $\{104\}$  plane is the most intense. In contrast, the number of peaks of original prism shell (Figure 2b) is much less than the powder XRD, indicating the shell has a preferred orientation, which has been confirmed by others. Dissolution obtained polymer also showed three broad peaks (Figure 2c) with d-space values of 1.568 nm ( $2\theta=5.633^\circ$ ), 0.3483 nm ( $2\theta=25.551^\circ$ ) and 0.2128 nm ( $2\theta=42.449^\circ$ ), implying these insoluble proteins are crystalline and might be ordered.

To prepare the sample for TEM observation, a piece of prismatic fragment was cut perpendicularly to organic matrix using FIB instrument. FIB is an ideal tool for TEM sample preparation with little artefact and is being widely employed in dealing with geomaterials and biominerals [41]. The cutting thin slice (Figure 3a) includes three different parts: two inorganic phases (on both sides) which come from two neighbouring prisms and an organic phase (middle part) which derives from the inter-prismatic matrix. The width of cutting thin slice is 15  $\mu\text{m}$ , while the thickness is less than 100 nm. The higher magnification of one of regions (circles) is shown in Figure 3b. One can observe that there are many black spots with average size of 3 nm inside the polymer, indicating the occurrence of different materials. Selected area electron diffraction (SAED) shows not only a ringed but also a spot pattern (Inset, Figure 3b), which indicates a preferred orientation for these nanocrystals. HRTEM image shows that the inter-atomic distances of these spots are 0.218 and 0.217 nm (Figure 3c), which correspond to  $\{202\}$  planes of the calcite crystal structure. Similarly, Velázquez-Castillo et al. [42] observed that nano-metric aragonite crystals embedded on the proteinous material of both matrix and the bridges in the *Nautilus pompilius*, which means that two species might adapt to same design strategies to complete mineralization.

Earlier studies showed that inter-crystalline macromolecules extracted from mollusk shell (both prismatic layer and nacreous layer) can affect the

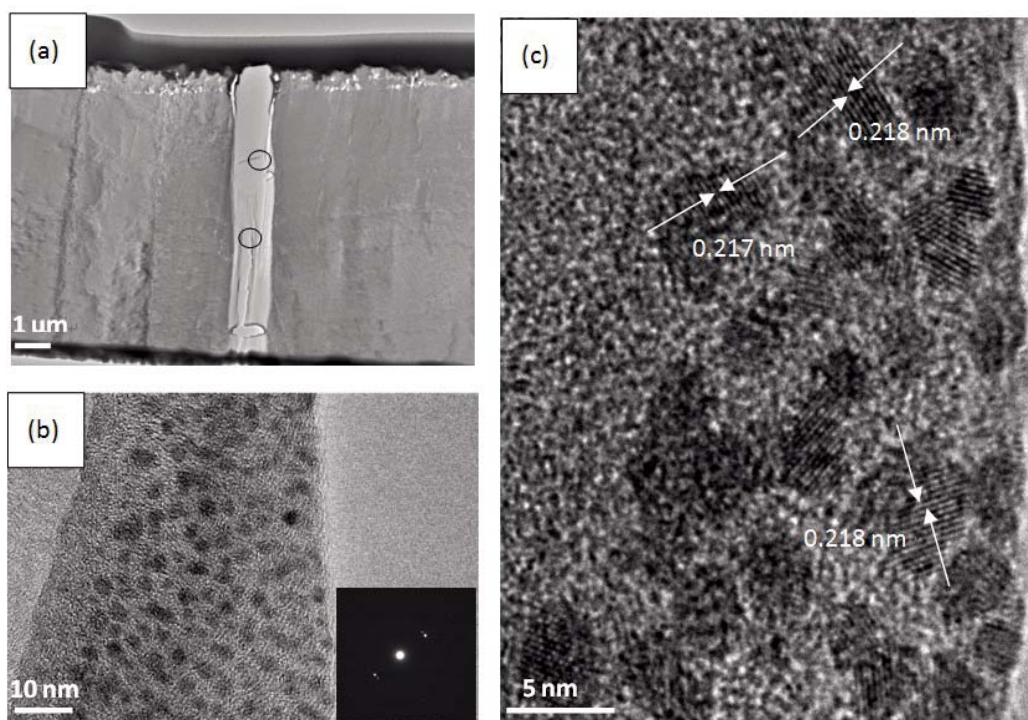


**Figure 2** Comparative X-ray diffraction patterns of (a) ground prism powder; (b) original prism outer surface; (c) polymer obtained by complete dissolution of inorganic phase.

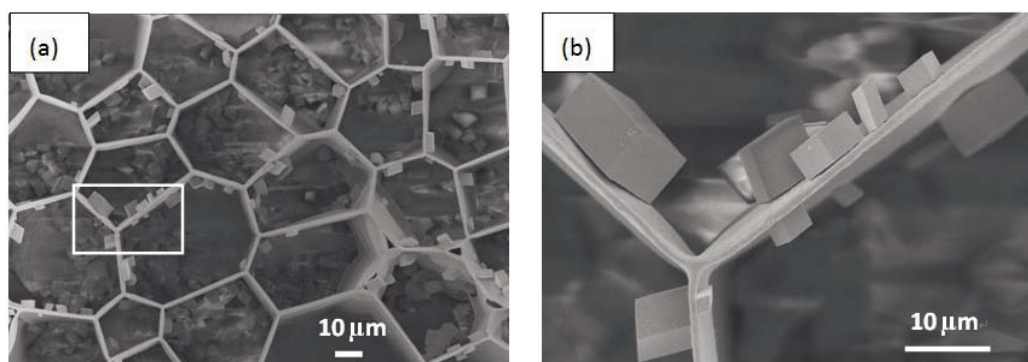
crystal polymorph and shape [24]. In order to study the effects of the insoluble intra-prismatic matrix to crystal nucleation, growth and orientation, we carried out  $\text{CaCO}_3$  crystallization experiments in vitro using HCl-partially dissolution obtained prism as template. After 6 hours mineralization time, some crystallites with the typical rhombohedral shapes are observed in Figure 4a. X-ray diffraction experiments (data not shown) confirm the presence of calcite phase of  $\text{CaCO}_3$ , which is consistent with the results of soluble proteins as additives from the same layer. Interestingly, some crystals adhered to the walls of organic matrix with certain  $\{104\}$  faces (figure 4b), while some regular calcite crystallites randomly oriented at the bottom. The preferential nucleation of  $\{104\}$  face indicates that macromolecules lateral walls contain appropriate chemical groups ( $-\text{COOH}$ ,  $-\text{OH}$ ) that interact with Ca ions in structural arrangements that resemble the ordering of the same ions in calcite particular face. In this way, the matrix acts as an organic template for epitaxial nucleation of  $\text{CaCO}_3$  through interfacial molecular recognition [43]. Since the lateral walls of inter-prismatic matrix could affect the crystallographic orientation in vitro experiment, we infer that it might act as the same role during mollusk shell formation. In fact, through the TEM observation on a single prism, we found that there are two differently oriented crystal arrangements which distribute in outside and inside, respectively (related data will publish elsewhere). This could be the results exerted together by inter- and intra-prismatic matrixes.

The fabrication of highly ordered mollusk shell is a rather complex process in which cells, macromolecules and ions are actively involved [38,44]. Epithelial cells in mollusk secrete organic fluids which self-organize into polygonal cavities by interfacial tension [18]. It is believed that the polygonal cavities are pre-formed before filling the  $\text{CaCO}_3$  inorganic phase. The inter-prismatic matrix is composed mainly of glycine-rich proteins [4,20]. According to Addadi et al.'s proposal [20], the growth mechanism of the prismatic layer in *Atrina rigida*





**Figure 3** (a) TEM image of a piece of foil cut perpendicularly to inter-prismatic matrix by FIB. The middle part is inter-prismatic matrix, which could be easily identified with both sides (inorganic phase) due to contrast. (b) magnified image of one of areas (circles) observed in (a). Inset is corresponding SAED pattern. (c) HRTEM image of one of areas in (b).



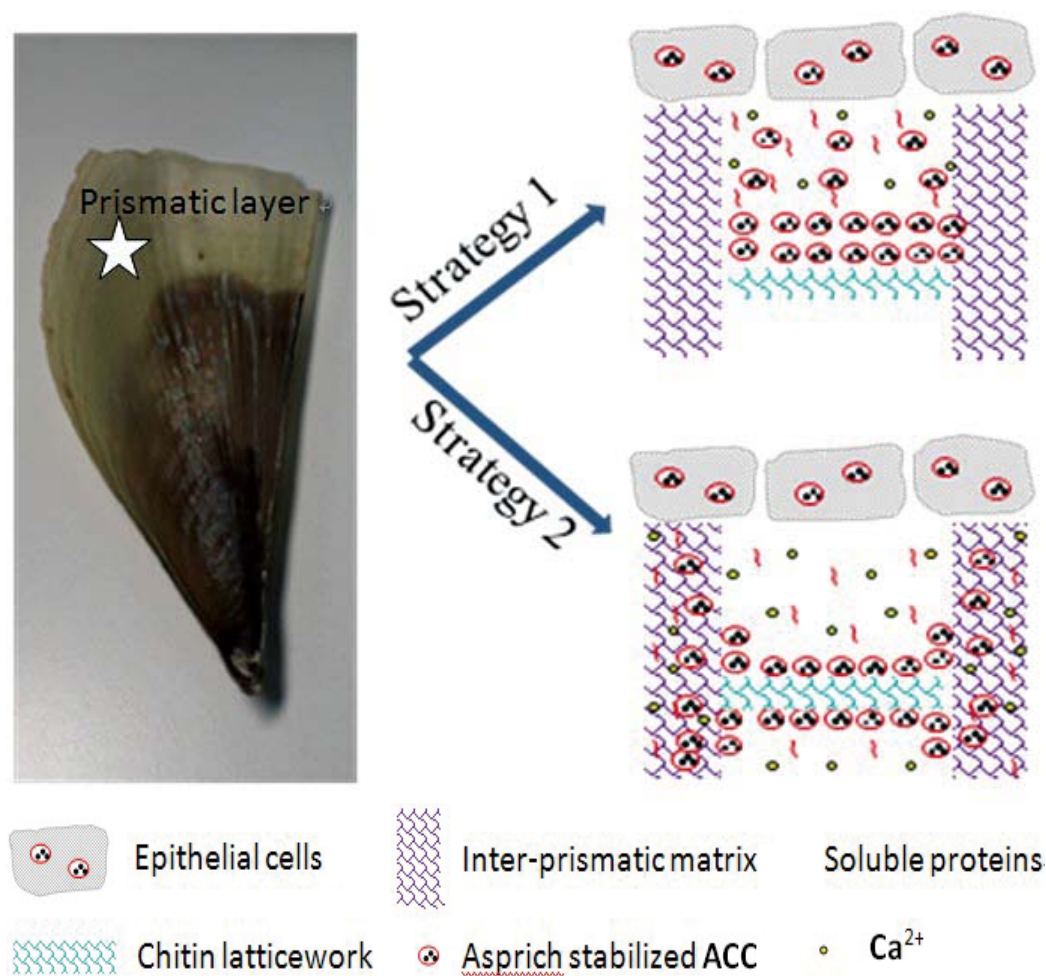
**Figure 4** SEM images of insoluble polymer as template directing the crystal growth. (a) the distribution of  $\text{CaCO}_3$  crystals on/in polymer. Some crystals with  $\{104\}$  faces are closely adhered to walls of inter-prismatic matrix, others randomly arranged at the bottom. (b) a higher amplification of rectangular area in figure 4(a). One could clearly see that the  $\{104\}$  faces of rhombohedral  $\text{CaCO}_3$  micro-crystallites are oriented parallel to the inter-prismatic walls.

basically adopted the way of layer by layer superposition with alternating layers of Asprich-stabilized amorphous calcium carbonate (ACC) and chitin (Figure 5, Strategy 1). Especially, the successful introduction of ACC could effectively resolve a list of paradoxes encountered in mollusk biomineralization [45]. However, based on their model, it is difficult for us to imagine how the prismatic layer which is closely adherent to the nacreous layer becomes thicker and thicker with the increments of age, as the mantle cells cannot make contact with this layer. On the base of our results, one alternative assumption is that there might exist a channel in the inter-prismatic matrix so that a part of calcium ions or Asprich stabilized ACC could pass through it to reach mineralization sites (Figure 5, Strategy 2). Meanwhile, these Asprich-stabilized ACC would fuse and crystallize inside the inter-prismatic matrix and on chitin latticework, respectively, due to the structural control. In the end, a monocrystalline

was formed with likely occluded macromolecules. We are inclined to think that Strategy 1 collaborates with Strategy 2 during the whole process. Additionally, the inter-prismatic matrix not only plays the function as framework but also affords some charged or polar residues as template for calcite oriented nucleation. In fact, Marin et al. accurately mapped the localization of casparin (a kind of acidic protein, from prismatic layer) by immunogold and discussed its different functions [17]. They pointed out that caspartin may be involved in maintaining the crystallographic orientation of the whole prism. Our in vitro recrystallization experiment unambiguously elucidates this point.

#### 4. Conclusion

In summary, we used dissolution and in situ recrystallization methods combining with modern materials



**Figure 5** Schematic representation of growth mechanism from prismatic layer. We briefly think that the complex process is a combination of two pathways. Strategy 1: layer by layer additions with alternating ACC and chitin fibers to achieve mineralization process. Strategy 2: ions, molecules and/or other forms of minerals through the channel of inter-prismatic matrix to complete mineralization process. See the text for detailed illustrations.

analysis and characterization techniques to fully unveil the structure information of inter-prismatic matrix in mollusk shell. A basic fact is that the Ca is included in the inter-prismatic matrix of formed shell. There could be a channel in the inter-prismatic matrix for the transport of ions, macromolecules and/or other forms of minerals. Through dissolution, porous and layer by layer structures are visible on inter-prismatic matrix. The walls of inter-prismatic matrix influenced the orientation of  $\text{CaCO}_3$  crystals in vitro. Our finding and “channel hypotheses” would enrich people’s understanding to mollusk biomineralization process. Further insights into the inter-prismatic matrix will be required to test our hypotheses.

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