

# Silver Nanoparticles: Novel Antimicrobial Agent Synthesized from an Endophytic Fungus *Pestalotia* sp. Isolated from Leaves of *Syzygium cumini* (L)

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## Abstract

We report the extracellular synthesis of silver nanoparticles using an endophytic fungus *Pestalotia* sp. isolated from leaves of *Syzygium cumini* (L) and their antibacterial activity against human pathogenic bacteria viz. *Staphylococcus aureus* (ATCC-25923) and *Salmonella typhi* (ATCC-51812) alone and in combination with commercially available antibiotics. Detection of synthesized silver nanoparticles was carried out using UV-Visible spectrophotometer analysis, which showed a peak at 415 nm indicating the formation of nanoparticles. Further characterization includes the Fourier Transform Infra-Red spectroscopic analysis for the detection of proteins as capping agents on nanoparticles. Nanoparticle Tracking and analysis (LM 20) and TEM analysis confirmed the formation of spherical and polydispersed nanoparticles in the range of 10-40 nm having average size of 12.40 nm.

Biologically synthesized silver nanoparticles showed significant antibacterial activity but their efficacy was increased in combination of antibiotics like gentamycin and sulphamethizole. Silver nanoparticles in combination with gentamycin showed maximum activity (30 mm) against *S. aureus* followed by sulphamethizole (25 mm). Similar results were reported in case of *S. typhi* where silver nanoparticles in combination with gentamycin (28 mm) showed more activity than combination of silver nanoparticles and sulphamethizole (24 mm).

Biosynthetic approach using an endophytic fungus is a novel way towards the development of safe, economically viable and green method for the synthesis of silver nanoparticles and thus synthesized silver nanoparticles can be used in antibacterial formulations.

**Keywords:** Endophyte, Silver Nanoparticles, Antibiotics, Economically viable, Synergistic effect

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## 1. Introduction

Metal nanoparticles have promising applications in the fields of medicine, electronics, agriculture, etc. In the present scenario pharmaceutical and biomedical sector are facing the challenge of continuous increase in the emerging pathogens, with their antibiotic resistance profiles, with fear about the emergence and re-emergence of multi-drug resistant pathogens and parasites [1]. Therefore, in this modern era the priority areas of research are concerning the development or modification in antimicrobial compounds in order to improve bactericidal potential.

Nanotechnology is the engineering and technological applications of the nanomaterials and nanoparticles of size ranging from (1-100 nm) [2, 3]. Nanotechnology provide platform to modify and develop the important

properties of metal in the form of nanoparticles having promising applications in diagnostics, biomarkers, cell labeling, contrast agents for biological imaging, antimicrobial agents, drug delivery systems and nano-drugs for treatment of various diseases [4, 5].

The researchers are moving towards nanoparticles especially silver nanoparticles to solve the problem of emerging pathogens including multi-drug resistant bacteria [6, 7]. Silver nanoparticles are more effective because of the high surface area to volume ratio so that a large proportion of silver nanoparticles is in direct contact with their environment [8].

Many fungi like *Fusarium acuminatum* [8], *F. solani* [9] *Aspergillus niger* [10] *Phoma glomerata*

[3] *Alternaria alternata* [11], *F. culmorum* [12] etc. have been successfully used for the synthesis of silver nanoparticles. These studies confirmed that among the different biological agents, fungi are more efficient candidates for fabrication of metal nanoparticles both intra- and extracellularly [13]. Extracellular biosynthesis of silver nanoparticles using fungi has advantages like more simple and eco-friendly approach as compared to chemical and physical methods [8]. The antimicrobial potential of silver nanoparticles have been examined and found to be effective against many pathogens [14, 15] Ingle et al [8]. demonstrated antibacterial activity of silver nanoparticles synthesized from *F. acuminatum* against human pathogenic bacteria like *S. typhi*, *E. coli*, *S. epidermidis* and multi-drug resistant *S. aureus*. Similarly, Gade and colleagues [10], reported antibacterial activity of silver nanoparticles against Gram-positive (*S. aureus*) and Gram-negative (*E. coli*) bacteria. Besides antimicrobial agents, silver nanoparticles are used in bio-labeling biosensors and filters [17], nano dressings and textile fabrics beneficial for the burnt patients, [18] for surgical masks, [19] in tissue conditioner [20], etc. Thus, silver nanoparticles are the ideal candidate for the development of novel antimicrobial product and these are said to be antimicrobials of new generations [21].

In the present study, we have used an endophytic *Pestalotia* sp. isolated from healthy leaves of *Syzygium cumini* (L.) for the extracellular synthesis of silver nanoparticles. We also evaluated their antibacterial activity. The main reason behind the use of endophytic fungus was that being an endophyte it is non pathogenic in nature and therefore, easy to handle and culture.

## 2. Materials and methods

### 2.1 Isolation and identification of endophytic *Pestalotia* sp.

Healthy leaves of *Syzygium cumini* (L) were collected from the campus of S. G. B. Amravati University, Amravati. Leaves were thoroughly washed in sterile distilled water and cut into small pieces. These pieces were surface sterilized using 1% NaOCl for 30 sec., washed with sterilized distilled water (2 times) and dried on paper towel. These pieces were then inoculated on petri-plates containing sterilized potato dextrose agar (PDA) and incubated at  $28 \pm 2^\circ\text{C}$  for 8-15 days. The fungus *Pestalotia* sp. have been identified on the basis of morphological and cultural characteristics like color of colony, growth rate, type of mycelium etc.

### 2.2 Extracellular synthesis of silver nanoparticles

For the synthesis of silver nanoparticles, the endophytic fungus *Pestalotia* sp. was grown in 250 ml flask containing 100 ml potato dextrose broth (PDB) at  $28^\circ\text{C}$  for 72 hours and then harvested biomass was filtered through Whatman filter paper No.1. The fungal mat was then washed with distilled water to remove media

component and suspended in 100 ml distilled water for 48 hours. After 48 hours of incubation, the cell filtrate was separated by filtration. Fungal cell filtrate was collected and challenged with the  $\text{AgNO}_3$  salt (final conc. 1 mM).

### 2.3 Detection of silver nanoparticles by UV-Visible spectrophotometer

After 2 hours of incubation, the reaction mixture was subjected to UV-Visible spectrophotometer analysis (Shimadzu UV-1700, Japan). The spectrum was scanned at the resolution of 1 nm from 200- 800 nm for each sample.

### 2.4 Characterization of silver nanoparticles

#### 2.4.1 Fourier Transform Infrared Spectroscopic (FTIR) Analysis:

After complete reduction of aqueous silver ions in to silver nanoparticle, the colloidal solution with acetone in (1:5) was subjected to centrifugation at 4000 rpm for 15 min after continuous shaking (2 times). Later, supernatant was discarded and 1-2 ml acetone was added into the pellet. After shaking thoroughly, it was poured into the petri plate. Acetone was then allowed to evaporate in order to obtain the powder of nanoparticles. Characterization of AgNPs was carried out by FTIR (Perkin- Elmer FTIR-1600, USA) in the range  $500\text{--}4000\text{cm}^{-1}$  at a resolution of  $4\text{cm}^{-1}$ . FTIR reveals the biomolecules responsible for the reduction of silver ions and stabilization of silver nanoparticles in the solution.

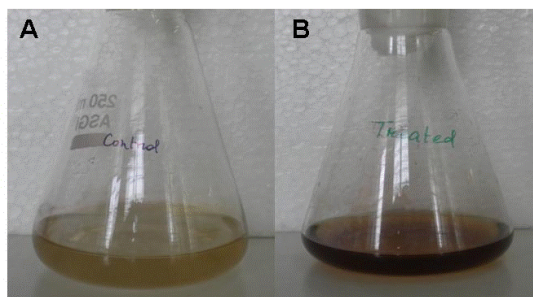
#### 2.4.2 Nanoparticle Tracking and Analysis System (NTA) (LM 20):

The size of synthesized nanoparticles was measured by Nanosight (LM-20, UK). The sample was diluted with the nuclease free water and 0.5 ml of diluted sample was injected onto the sample chamber and observed through LM 20 to measure the size of the nanoparticles.

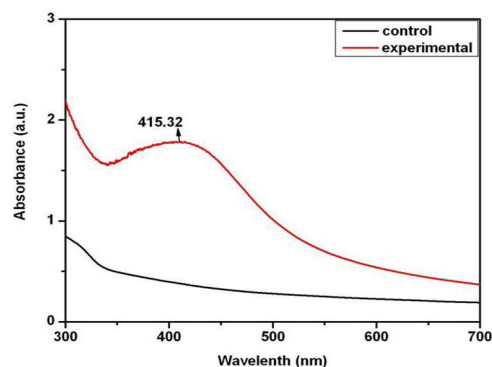
**2.4.3 TEM analysis:** The characterization of silver nanoparticles was also done by TEM (Philips, CM 12), on conventional carbon coated copper grids (400 meshes, Plano GmbH, Germany). A 5  $\mu\text{l}$  of sample was taken for the characterization and three image of each sample were taken for the clarification of the composition

### 2.5 Evaluation of antibacterial activity of silver nanoparticles

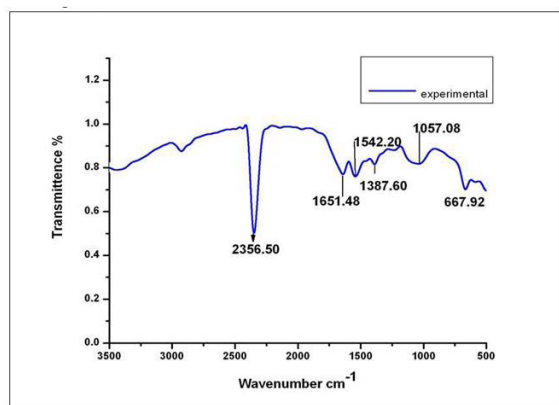
Antibacterial activity of silver nanoparticles alone and in combination with antibiotics was evaluated using Kierby-Bauer disc diffusion method, against *Staphylococcus aureus* (ATCC-25923) and *Salmonella typhi* (ATCC-51812) procured from American Type Culture Collection Center, USA. The Mueller- Hinton agar medium was prepared and sterilized at  $121^\circ\text{C}$  and 15 lbs pressure for 15 min, poured and allowed to solidify in the sterilized petri-plates. The overnight grown bacterial cultures of both bacteria having approx.  $10^5\text{ CFU ml}^{-1}$  were inoculated; four different types of disks control, silver nanoparticle, standard antibiotic Gentamycin and



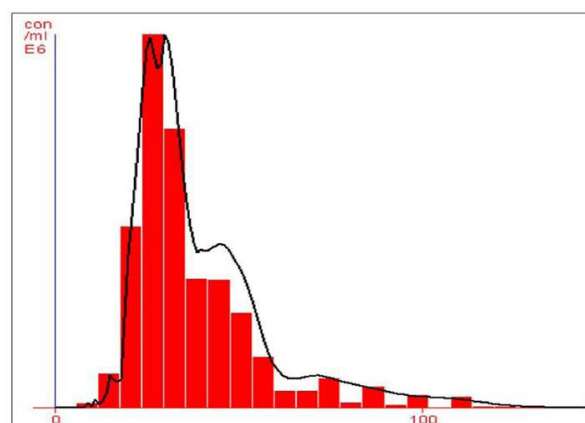
**Fig. 1** Synthesis of silver nanoparticles from endophytic *Pestalotia* sp. (A) Fungal cell filtrate before treatment of  $\text{AgNO}_3$  (control) and (B) Fungal cell filtrate after treatment of  $\text{AgNO}_3$  (experimental).



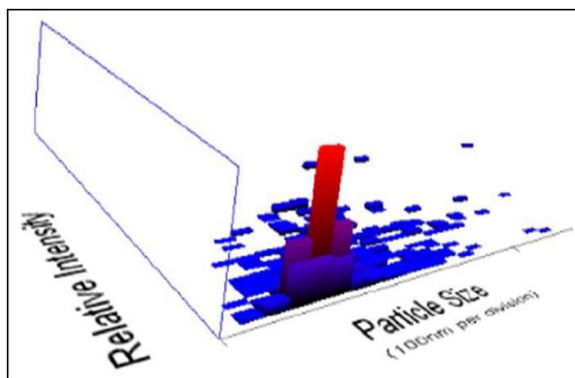
**Fig. 2** UV-Visible spectra of fungal cell filtrate (showing no peak) and silver nanoparticles showing peak at 415 nm.



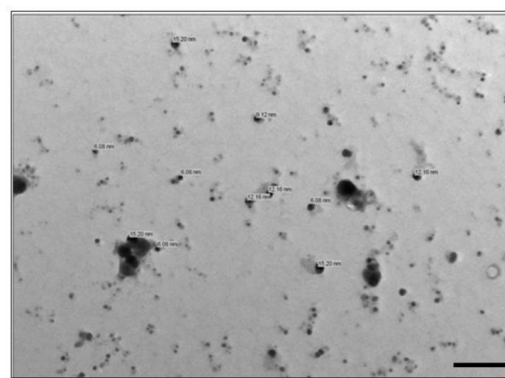
**Fig. 3** FTIR spectrum showing the presence of proteins as capping agent for silver nanoparticles.



**Fig. 4** NTA (Nanosight-LM 20) nanoparticle size distribution histogram showing the average size of 26 nm.



**Fig. 5** NTA (Nanosight-LM 20) 3-D plot of nanoparticle size distribution.



**Fig. 6** TEM micrograph showing synthesis of polydispersed and spherical silver nanoparticle having average size of 12.40 nm (scale bar 100 nm).

**Table 1.** Comparison of antibacterial activity of silver nanoparticles, synthesized from endophytic fungus *Pestalotia* sp, alone and in combination with antibiotics against test bacteria.

Antibiotics	Test Bacteria	Zone of Inhibitions (Antibiotic) in mm (A)*	Zone of Inhibition (Nanoparticles) in mm	Zone of Inhibitions (Antibiotic + Silver nanoparticle) in mm (B)*	Increase in Fold Area of Zone of Inhibition in mm $+(B^2 - A^2)/A^2$
Gentamycin	<i>S. aureus</i>	27	12	30	0.23
	<i>S. typhi</i>	26	10	28	0.15
Sulphamethizole	<i>S. aureus</i>	23	12	25	0.18
	<i>S. typhi</i>	23	10	24	0.08

\*Mean surface area of the inhibition zone was calculated for each from the mean diameter. +Increase in fold area was calculated as  $(B^2 - A^2)/A^2$ , where 'A' and 'B' are the inhibition zones for Antibiotic and Antibiotic + silver nanoparticles respectively.



sulphamethizole purchased from Hi-Media (Mumbai, India) and the combination of silver nanoparticles with standard antibiotic mentioned above were placed on the surface of the agar. Plates were incubated for 24 hours at 37°C, Zone of inhibition was measured. The assay was performed in triplicates.

## 2.6 Assessment of increase in fold area

The increase in fold area was assessed by calculating the mean surface area of the inhibition zone of each antibiotic (gentamycin and sulphmethizole) and antibiotic + silver nanoparticle. The fold increase area of *S. aureus* and *S. typhi* was calculated by the equation  $(B^2 - A^2)/A^2$  where 'A' and 'B' were zones of inhibition for antibiotic and antibiotic + silver nanoparticles, respectively [3].

## 3. Results and Discussion

The rapid change in color from yellowish to dark-brown was observed in the fungal cell filtrate after addition of the aqueous silver ions ( $\text{AgNO}_3$ ) due to the reduction of silver ions to silver nanoparticles i.e.  $\text{Ag}^+$  to  $\text{Ag}^0$ . The appearance of brown color indicates the synthesis of silver nanoparticles (Fig. 1). These findings showed the similarity with the results reported by many researchers in past [8, 22, 23], who used different fungal system for the synthesis of silver nanoparticles. The formation of silver nanoparticles in fungal cell filtrate was further characterized by using UV-Vis spectrophotometer based on its characteristics surface plasmon resonance [8]. The reaction mixture after treatment with aqueous silver ions and subjected to optical analysis using UV-Vis spectrophotometer showed a sharp peak at 415 nm which is specific for silver nanoparticles and appeared due to plasmon resonance [24] (Fig. 2).

FTIR spectrum revealed that the silver nanoparticles synthesized from endophytic fungus *Pestalotia* sp. showed the presence of peak at 1651.4, 1542, 1387 and 1057  $\text{cm}^{-1}$  (Fig. 3). The bands at 1651.4, 1542.4 corresponds to the bonding vibrations of the amide I and amide II bands of proteins while the bands obtained at 1387  $\text{cm}^{-1}$  and 1057  $\text{cm}^{-1}$  are due to presence of C-N stretching vibrations of aromatic and aliphatic amines. These results resembles with the findings of Gole and colleague [25] who reported that proteins can bind to nanoparticles either through free amine groups or cystein residues or through the electrostatic attraction of negatively charged carboxylate groups in enzymes present in cell-wall of mycelia. Non-conjugated aliphatic C=C and C=N have absorption bands at 1690 to 1620  $\text{cm}^{-1}$ , with variable intensities. Molecules containing  $\text{NO}_2$  groups, such as nitro compounds, nitrates, and nitramines, commonly exhibit asymmetric and symmetric stretching vibrations of the  $\text{NO}_2$  group at 1660 to 1500 and 1390 to 1260  $\text{cm}^{-1}$  region. FTIR spectroscopy has confirmed that amino acid residues and peptides of proteins has the stronger ability to bind with metal, so that the proteins could most possibly form a coat covering the metal nanoparticles i.e. (capping of silver nanoparticles) to prevent agglomeration of the particles and stabilizing

in the medium [26].

Nanoparticle tracking and analysis (NTA) was carried using NanoSight LM-20 to measure the dispersion characteristics, i.e. size and distribution. In particular, it is the most recently developed system; NTA was assessed in-depth due to its ability to measure the size of particles individually on a particle-by-particle basis. NTA allows individual nanoparticles in a suspension to be microscopically visualized and their Brownian motion to be separately but simultaneously analyzed, from which the particle size distribution can be obtained. The average size of nanoparticles from the NTA analysis was found to be 26 nm. Fig. 4 and Fig. 5 showed the particle size distribution histogram and 3-D plot of particle size distribution respectively. These results corroborate the results obtained by Montes-Burgos and group [27]. TEM analysis finally confirmed the synthesis of spherical and polydisperse silver nanoparticles in the reaction mixture. The particles are in the range of 10-40 nm with average diameter of 12.40 nm (Fig. 6).

*In vitro* antimicrobial activity, of silver nanoparticles produced from endophytic fungus *Pestalotia* sp. was carried out without antibiotics and in combination with commercially available antibiotics gentamycin and sulphamethizole against *S. aureus* and *S. typhi*. Silver nanoparticles without antibiotics showed antibacterial activity but the efficiency was found to be increased significantly in combination with antibiotics. Silver nanoparticles in combination with gentamycin showed maximum activity (30 mm) (increase in fold area-0.23) against *S. aureus* followed by sulphamethizole (25 mm) (increase in fold area-0.18). Similar results were reported in case of *S. typhi* where silver nanoparticles in combination with gentamycin (28 mm) (increase in fold area-0.15) showed more activity than combination of silver nanoparticles and sulphamethizole (24 mm) (increase in fold area-0.08) (Table 1).

Above results corroborates with the findings reported by Birla et al [3]. who reported that *in vitro* activity of commercially available antibiotics was significantly increased in the presence of silver nanoparticles produced from *Phoma glomerata*. Similarly, Gajbhiye et al [11] also demonstrated the increase in the efficacy of silver nanoparticles in presence of standard antibiotics.

## 4. Conclusion

It has been demonstrated that the endophytic fungus *Pestalotia* sp. is capable of producing silver nanoparticles extracellularly which are quite stable in solution due to capping by the proteins present in the cell filtrate. This is an efficient, eco-friendly and simple process. The silver nanoparticles showed significant antibacterial activity. Their efficacy increased when used in combination with commercially available antibiotics. Therefore, such silver nanoparticles can be used as antimicrobial agent alone or in combination with antibiotics after further trials on experimental animals.

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